Accepted Manuscript

Title: Delayed Acquisition of Neonatal Reflexes in newborn Primates receiving A Thimerosal-containing HepatitiS B Vaccine: influence of gestational age and Birth weight

Authors: Laura Hewitson, Lisa A. Houser, Carol Stott, Gene Sackett, Jaime L. Tomko, David Atwood, Lisa Blue, E. Railey White, Andrew J. Wakefield



PII:	S0161-813X(09)00222-8
DOI:	doi:10.1016/j.neuro.2009.09.008
Reference:	NEUTOX 1068
To appear in:	NEUTOX
Received date:	16-6-2009
Revised date:	9-9-2009
Accepted date:	17-9-2009

Please cite this article as: Hewitson L, Houser LA, Stott C, Sackett G, Tomko JL, Atwood D, Blue L, White ER, Wakefield AJ, Delayed Acquisition of Neonatal Reflexes in newborn Primates receiving A Thimerosal-containing HepatitiS B Vaccine: influence of gestational age and Birth weight, *Neurotoxicology* (2008), doi:10.1016/j.neuro.2009.09.008

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

*Manuscript

ACCEPTED MANUSCRIPT

DELAYED ACQUISITION OF NEONATAL REFLEXES IN NEWBORN PRIMATES RECEIVING A THIMEROSAL-CONTAINING HEPATITIS B VACCINE: INFLUENCE OF GESTATIONAL AGE AND BIRTH WEIGHT

- 4 *Laura Hewitson^{1,3}, Lisa A. Houser¹, Carol Stott³, Gene Sackett², Jaime L. Tomko¹, David Atwood⁴,
- 5 Lisa Blue⁴, E. Railey White⁴ and Andrew J. Wakefield³
- 6 ¹Department of Obstetrics and Gynecology, University of Pittsburgh School of Medicine,
- 7 Pittsburgh, PA 15213, ²Washington National Primate Research Center, University of Washington,
- 8 Seattle, WA 98195, ³Thoughtful House Center for Children, Austin, TX, 78746 and ⁴Department of
- 9 Chemistry, University of Kentucky, Lexington, KY 40506
- 10
- 11 *Corresponding Author:
- 12 Laura Hewitson, Ph.D.
- 13 Magee-Women's Research Institute and Foundation
- 14 University of Pittsburgh School of Medicine
- 15 204 Craft Avenue, Pittsburgh, PA 15213
- 16 <u>LHewitson@mwri.magee.edu</u>
- 17 412-641-2490 (Ph); 412-641-2410 (Fax)
- 18

1 Abstract

2 This study examined whether acquisition of neonatal reflexes and sensorimotor skills in newborn 3 rhesus macaques (Macaca mulatta) is influenced by receipt of the single neonatal dose of Hepatitis B 4 (HB) vaccine containing the preservative thimerosal (Th). HB vaccine containing a standardized 5 weight-adjusted Th dose was administered to male macaques within 24 hours of birth (n=13). 6 Unexposed animals received saline placebo (n=4) or no injection (n=3). Infants were raised identically 7 and tested daily for acquisition of 9 survival, motor, and sensorimotor reflexes by a blinded observer. 8 In exposed animals there was a significant delay in the acquisition of three survival reflexes: root, snout 9 and suck, compared with unexposed animals. No neonatal responses were significantly delayed in 10 unexposed animals compared with exposed. Gestational age (GA) and birth weight were not 11 significantly correlated. Cox regression models were used to evaluate the main effects and interactions 12 of exposure with birth weight and GA as independent predictors and time-invariant covariates. 13 Significant main effects remained for exposure on root and suck when controlling for GA and birth 14 weight such that exposed animals were relatively delayed in time-to-criterion. There was a significant 15 effect of GA on visual follow far when controlling for exposure such that increasing GA was associated 16 with shorter time-to-criterion. Interaction models indicated that while there were no main effects of 17 GA or birth weight on root, suck or snout reflexes there were various interactions between exposure, 18 GA, and birth weight such that inclusion of the relevant interaction terms significantly improved 19 model fit. This, in turn, indicated important influences of birth weight and/or GA on the effect of 20 exposure which, in general, operated in a way that lower birth weight and/or lower GA exacerbated 21 the detrimental effect of vaccine exposure. This primate model provides a possible means of assessing 22 adverse neurodevelopmental outcomes from neonatal Th-containing HB vaccine exposure, 23 particularly in infants of lower GA or low birth weight. The mechanism of these effects and the 24 requirements for Th is not known and requires further study.

- Key Words: Macaca mulatta; animal model; Hepatitis B vaccine; ethyl mercury; thimerosal;
 neurodevelopment; neurotoxicity.
- 3

1 1. Introduction

The Hepatitis B (HB) vaccine was introduced into the US childhood immunization schedule in 1991 [1]. This schedule recommended that all infants irrespective of gestational age (GA) and birth weight born to HB-negative mothers be immunized with a HB vaccine within 12 hours of birth (i.e. before hospital discharge [1, 2]. We were unable to identify preclinical or prospective neurotoxicity studies that assessed the safety of this policy.

7

8 The two formulations of HB vaccine manufactured during the 1990's contained the preservative 9 thimerosal (Th), an antibacterial and fungistatic agent composed of ethyl mercury and thiosalicylate. 10 The HB vaccine contained 12.5 micrograms (µg) ethyl mercury, given to neonates unadjusted for GA or birth weight. Following safety concerns, particularly in respect of the potential for 11 12 neurotoxicity, a Congressionally-mandated Food and Drug Administration (FDA) review in 1999 13 produced a recommendation for the reassessment of Th use in vaccines. By 2001 the majority of 14 pediatric vaccines routinely recommended in the U.S. for children 6 years of age and under were 15 produced without Th, with the exception of multi-dose inactivated influenza and meningococcal 16 polysaccharide vaccines.

17

Since Th-containing vaccines, including the neonatal HB vaccine, continue to be used routinely in developing countries [3], continued safety testing is important, particularly for premature and lowbirth-weight neonates. Several formulations of HB vaccine currently used in other countries contain 25-50 µg ethyl mercury [3]. Based on the type of HB vaccine administered and the baby's birth weight there can be a 10-fold difference between the highest and lowest levels of mercury exposure to neonates [3]. In a U.S. study performed prior to the removal of Th from HB vaccines, blood mercury levels were significantly elevated in both preterm and term infants post HB vaccination [4].

While blood mercury levels are a poor reflection of body-burden of mercury, it is notable that these levels were higher in the pre-term infants when compared with term infants [4]. These findings suggest that newborns, especially pre-term infants, might have decreased ability to eliminate mercury since hepatic metallothionein and glutathione synthesis, both requirements for efficient mercury elimination, are not present in the neonate [5, 6].

6

7 Here we examine, in a prospective, controlled, observer-blinded study, the development of neonatal 8 reflexes in infant rhesus macaques after a single dose of Th-containing HB vaccine given within 24 9 hours of birth, following the US childhood immunization schedule (1991-1999). The rhesus macaque is used in preclinical vaccine neurotoxicity testing and displays complex early 10 11 neurobehavioral and developmental processes that are well characterized (reviewed by [7]). Neonatal 12 motor reflexes, also referred to as 'survival' reflexes, are necessary for survival of macaques in the 13 natural environment [8]. If an infant primate is unable to suck, for example, they will not be able to 14 obtain milk and are at risk of starvation. Neurobehavioral tests to assess early neonatal behavioral 15 functioning are therefore commonly used to detect effects of post-natal events or interventions [9], 16 such as exposure to organomercurials [10]. Macaques have also been used extensively in previous 17 studies of methyl and ethyl mercury toxicokinetics and developmental neurotoxicity [11-16] making 18 them a preferred model for addressing possible neurodevelopmental concerns regarding vaccine 19 safety. Male primates were chosen because of the male preponderance of neurodevelopmental 20 disorders and mounting evidence in support of a gender-selective neurotoxicity of organomercurials 21 in both humans and animals [17-22]. The objective of this study was to examine, in a primate model, 22 whether administration of a single thimerosal-containing hepatitis B vaccine at birth - the 23 recommended pediatric protocol throughout the 1990's - resulted in delays in the acquisition of 24 neonatal reflexes.

1 2. Materials and Methods

2 2.1 Animal Assurances

All procedures used in this research followed the guidelines of the Animal Welfare Act and the
Guide for Care and Use of Laboratory Animals of the National Research Council. Research
protocols were approved by the University of Pittsburgh/MWRI&F Institutional Animal Care and
Use Committees (IACUCs).

7

8 2.2 Subjects

Twenty nursery-reared rhesus macaque infants served as study subjects at the Pittsburgh 9 10 Development Center primate nursery. Pregnancies were produced by time-mated breeding. Fertile 11 dams were selected based on their menses records and placed with a proven breeder for 4-5 days 12 starting 2 days prior to expected ovulation. Mating was indicated by the presence of a seminal plug 13 and GA confirmed by ultrasound at approximately 30 days. After delivery the health of infants was 14 assessed by Simian APGAR, vital signs, and physical appearance. The birth weight and GA of the 15 infant monkeys were within the normal range for this species; the average birth weight was 529g (SD 16 78.4g; range, 394 - 688g) and average GA was 168 days (SD 5.51 days; range, 157 - 178 days).

17

18 **2.3.** Housing

19 Infants were separated from their mothers at birth and reared within a neonatal nursery according to 20 the detailed protocols of Ruppenthal and Sackett [7]. Separation was necessary for this study as 21 mothering precludes neonatal testing due to the distress caused to both the mother and the neonate 22 when temporarily separated [23-24]. The only way to rigorously test neonatal behavioral 23 development is to remove the infant from its mother at birth [7, 23]. This protocol also ensures the 24 survival of infants irrespective of birth weight or GA [25]. Environmental conditions were strictly

1 controlled in the nursery to eliminate potential confounding factors such as diet or infant handling. 2 Infants were similarly isolated, housed, and bottle-fed by hand in the nursery until achieving 3 temperature regulation, typically 7-10 days from birth. For the first three days of life, vital statistics 4 (respiration, heart rate, and temperature) were taken every 4 hours. Infants that could self-regulate 5 temperature during this three-day period were moved out of their incubator and singly housed in a 6 small nursery cage with a heating pad. If infants remained stable from Days 4-10, the heating pad 7 was turned off. The cage had mesh walls on all sides to provide the infant with good visibility of his 8 environment and also contained a cylindrical shaped hanging cloth surrogate suspended from the 9 cage ceiling. Infants could see and hear each other but had no physical contact, both within and 10 between peer groups for the duration of this study as per standard nursery procedures. Each cage 11 also contained a formula feeder used to train infants to feed themselves. They received a standard 12 infant baby formula (Enfamil, Mead Johnson and Co., IN). The appetite, attitude, activity level, 13 hydration status, and stool quality of each animal were assessed by a nursery technician at each 14 feeding. If infants were able to maintain health with the heating pad off and were self feeding, their 15 heating pads were removed by Day 14. Lights were on in the nursery from 0600 h to 2000 h. Room 16 temperature was maintained between 75-77°F.

17

18 2.4 Study Design

Animals were allocated to either the vaccinated (exposed) or saline/no injection (unexposed) groups on a semi-random basis in order to complete peer groups for later social testing [7] such that each peer group contained animals from either the unexposed or exposed study groups. Once a new peer group was started, new animals were assigned to this group until it consisted of 3 or 4 infants, the ages of which were less than 4 weeks apart from their peers. Infants received either a single dose

1 (0.5ml) of Th-containing HB vaccine (n = 13) or a saline injection (n = 4) both administered i.m. in
2 the thigh within 24 hours of birth, or no injection (n = 3).

3

4 2.5 Vaccine Source, Preparation and Dosing

5 In July 1999, the CDC and American Academy of Pediatrics (AAP) recommended that thimerosal 6 (Th) should be removed from pediatric vaccines, including all Hepatitis B (HB) formulations, which 7 until that time contained 12.5 µg ethyl mercury per 0.5ml dose. Therefore, in order to recreate the 8 HB vaccine used in the 1990's, a Th-free preparation of this vaccine RECOMBIVAX HB® (Merck) 9 was purchased and Th added as described below. The amount of Th added to the HB vaccines 10 resulted in a concentration of 2.0 µg ethyl mercury per 0.5ml dose. This represents a 6.25 fold 11 reduction of ethyl mercury in macaque HB vaccines compared with those formulated for human 12 use. This was necessary to adjust for the smaller size of the rhesus infants [26], thereby maintaining a 13 similar ethyl mercury 'exposure' to human infants of $\sim 2 \mu g \text{ kg/body weight}$. Purchased HB vaccines 14 were pooled prior to Th addition. Th dosing and all QA/QC were performed at the University of 15 Kentucky in the Environmental Research and Training Laboratory (ERTL). Stock Th (Sigma-16 Aldrich, St. Louis, MO) solutions were prepared such that a 50 µl dose added to the pooled vaccines 17 would vield the desired ethyl mercury concentrations. Triplicate stock Th solutions and spiked 18 vaccine solutions were digested in 5% nitric acid at 100°C for two hours and analyzed for ethyl 19 mercury concentration using a Varian Vista Pro CCD Simultaneous Inductively Coupled Plasma Optical Emission Spectrometer (ICP-OES) to verify that target concentrations were achieved. 20 21 Matrix effects were evaluated and corrected for using an yttrium internal standard. Furthermore, 22 second source curve verifiers and spike recoveries were in excess of 95%. Laboratory Control 23 Samples (LCSs) consisting of three different dilutions of the stock solutions bracketing the expected 24 concentrations of the dosed vaccines were also prepared and analyzed alongside the dosed vaccines

- on a Nippon MA-2000 mercury analyzer. Recoveries on the LCSs were again in excess of 95%. The
 dosed HB vaccine contained ~2.0 µg ethyl mercury per 0.5ml dose.
- 3

4 2.6 Neurodevelopmental Testing

5 From birth, the development of neonatal and infant reflexes and perceptual and motor skills were 6 assessed in all infants [7]. These were based on the Brazleton assessment scale, which was originally 7 developed for human infants [27]. Tests were performed daily and measured basic motor reflexes, 8 visual and auditory orienting, muscle tone, and behavioral state as described below. Responses and 9 scoring criteria are described in Table 1 and have been extensively published [7-9, 28-30]. Neonatal 10 assessments were performed by L.A.H. who was unaware of the study group assignment of each 11 animal, the number of animals in each study group, and the number of study groups. L.A.H. 12 underwent extensive training by G. Ruppenthal, one of the co-authors of the 'Research Protocol and 13 Technician's Manual', the most exhaustive guide on the healthy development of nursery-raised 14 infant macaques [7]. Training of L.A.H. was completed on non-study infants prior to the acquisition 15 of data for this study and consisted of comparison of assessments of similarly aged infants collected 16 by multiple trained testers. Reliability for nursery assessments by the trainee was achieved when they 17 obtained an 89% agreement with the trained testers on three consecutive randomized assessments. 18 As part of standard protocol [7], nursery testers who worked in the facility were routinely retested 19 against each other at 3-6 months intervals.

20

All assessments of reflex acquisition were performed in a designated area under strictly controlled environmental conditions. Testing was always daily at a time intermediate between feedings when the neonate is least likely to be asleep or hungry [7]. For each test, the infant was removed from its cage, wrapped in a cloth diaper, and stimulated to perform the following ten behaviors: (i) four basic

1 'survival' reflexes including root (elicited by lightly brushing the cheek from ear to mouth), snout 2 (elicited by brushing downward from the forehead between the eyes to the tip of the nose), suck 3 (elicited by inserting a nipple into the mouth), and *auditory startle* (elicited by dropping a small metal 4 object behind the infant's head); (ii) two motor reflexes including grasp (elicited by placing a finger in 5 the palm of the hand and on the bottom of the foot), and *clasp* (elicited by placing the hands and feet 6 around a cloth-covered cylinder held above the infant; and (iii) three sensorimotor behaviors 7 including *auditory orienting* (elicited by lip smacking, a mother-to-infant communication gesture), visual 8 orienting and following (elicited by positioning a small toy in front of the infant's face then moving the 9 toy from left to right and right to left). The visual procedures were done with the toy positioned at 10 either 12 or 24 inches in front of the infant representing both near and far visual tests, respectively. 11 Although a total of 9 neonatal reflexes were measured, the inclusion of both hand/foot for grasp and 12 *clasp* and near/far for visual procedures resulted in a total of 13 variables analyzed. The criterion for each measure was reached once the infant displayed the highest possible score (see Table 1). 13

14

15 Insert Table 1.

16

17 2.7 Statistical Analyses

Analyses were carried out using SPSS v. 17 (SPSS, Inc., Chicago, IL). Analyses were based on the assessments of 20 animals from the day of birth to 14 days of age. The dependent variable in all survival analyses was hazard of time-to-criterion, measured for all neonatal reflexes and sensorimotor responses within this time frame. Right censoring of observations was applied when animals failed to reach criterion by Day 14. This was the required day of censoring since infants received further interventions on Day 14 which would have confounded the independent effects of the HB vaccine. Non-parametric Kaplan-Meier (K-M) log-rank estimation was used to estimate

differences in age at criterion for each neonatal response between exposed and unexposed animals.
 Differences in survival curves associated with exposure status were tested using the log-rank test.
 The effect of GA and birth weight as independent predictors and time-invariant covariates was
 examined using Cox's regression.

5

6 The K-M log-rank test is a non-parametric method for comparing the survival experience of two or 7 more groups, but it cannot be used to evaluate the effects of several variables on survival. The 8 regression method introduced by Cox ([31]; proportional hazards regression analysis) is widely used when there is a need to investigate several variables at the same time. Variables entered into the 9 10 model simultaneously generate output statistics which indicate the effect of each variable on 11 outcome, while controlling for all other variables included in the model. These are referred to as 12 'main effects'. As in all regression modeling it is important to evaluate both the 'main effects' of 13 variables and their possible interactions. Adding interaction terms allow exploration of the 14 potentially differential effect of a variable across different exposure groups – asking the question of 15 whether the effect of, in this case, either GA or birth weight, operates differently in exposed vs. 16 unexposed animals.

17

Statistical significance was set at p≤0.05 but variables approaching significance have also been noted where relevant. This is particularly important given the relatively low power of the study. Six variables included left-censored values for two unexposed animals who had reached criterion at first assessment; a score of 0.5 was allocated in each instance. For all regression models the Exp(β) variable is the main variable of interest indicating the predicted change in risk of meeting criterion (hazard) for a unit increase in the predictor.

24

1 When comparing models for goodness of fit (i.e how well the particular model accounts for the 2 outcome of interest) there are two relevant values. First, the likelihood-ratio test for the hypothesis 3 that all parameters are 0 (no independent effects) is obtained by comparing the log-likelihood (x -2; 4 -2LL) estimated for an Omnibus model in which all coefficients are 0, with the -2LL for the model 5 that contains all the variables of interest. The lower the -2LL, the better the model. The difference 6 between these models is also represented by the Chi-Square (χ^2) Change Statistic and its 7 accompanying p value. After adding terms to the model, if the observed significance level of the 'Model Change χ^2 ' is small (≤ 0.05) the null hypothesis, that all model coefficients are 0, can be 8 rejected. None of the Cox Regression models violated the proportional hazards assumption which 9 10 requires that the hazard ratio (hazard in exposed/hazard in unexposed) is constant over time.

11

12 **3. Results**

13 **3.1 Infant Health**

14 All infants remained healthy during the study testing period reaching all criteria for maintaining 15 health including appetite, weight gain, and activity level, and achieved temperature regulation by Day 3. GA fell within normal range for all animals and was distributed normally in the sample, ranging 16 17 from 160 to 178 in exposed animals (mean 169.08, SD 5.31) and from 157 to 176 in unexposed 18 animals (mean 167.57, SD 6.16). Birth weight also fell within normal range for all animals. 19 Distribution of birth weight was slightly skewed to the right in the sample, ranging from 394 to 688 20 in exposed animals (mean 524.40, SD 77.51; median 502, IOR 95.50) and from 449 to 643 in 21 unexposed animals (mean 538, SD 85.54; median 494, IQR 171). There were no statistically 22 significant differences between exposed and unexposed animals in either GA or birth weight (p>0.5) 23 and no statistically significant correlation between GA and birth weight either for the sample as a 24 whole, or for exposed vs. unexposed animals (p > 0.5).

1 3.1 Neonatal Development

2 Neonatal reflexes and sensorimotor responses were measured daily from birth until post-natal Day 3 14. Datasets from the two unexposed groups (with or without a saline injection) were combined 4 when no differences were found for all measures (p>0.5). There was a significant delay in time-to-5 criterion for exposed vs. unexposed animals for three survival reflexes including root (Fig. 1A; 6 p=0.004), suck (Fig. 1B; p=0.002) and snout (Fig. 1C; p=0.03) and approached significance for startle 7 (p=0.11). The effect of exposure also approached significance for grasp hand (p=0.07), one of the 8 motor reflexes. There were no reflexes for which the unexposed animals took significantly longer to 9 reach criterion than the exposed animals (Table 2).

10

11 Insert Figure 1 and Table 2

12

13 **3.2 Modeling Time-to-Criterion: Main Effects Models**

Further evaluation of the potential impact of GA and/or birth weight on the association between exposure and outcome was undertaken using Cox regression models. Before including potential covariates, unadjusted main effects of exposure were evaluated and confirmed for *root* (Exp(β)=3.95, 95% CI 1.29 – 12.05, *p*=0.018) and *suck* (Exp(β)=4.58; 95% CI 1.41 – 14.93; *p*=0.011). For *snout* the effect of exposure did not maintain its significance in the model (Exp(β)=3.18; 95% CI 0.952 – 10.615; *p*=0.06).

20

3.2.1 Exposure and Gestational Age (GA): Cox regression modeling for main effects of exposure and GA together demonstrated a significant main effect of exposure for *root* (p=0.009) and *suck* (p=0.015) such that unexposed animals reached criterion more quickly. This approached statistical significance for *snout* (p=0.055). There was no main effect of GA for *root, suck* or *snout* (p>0.05).

When GA was included in the model, a significant main effect of GA on *visual follow far* was
 observed (*p*=0.013) such that an increase in GA was associated with animals reaching criterion more
 quickly (Table 3). The unadjusted effect of GA on *visual follow far* was not statistically significant.

4

5 3.2.2 Exposure and Birth Weight. Cox regression modeling of exposure and birth weight 6 demonstrated significant main effects that were almost identical to the models for exposure and GA 7 described above. There were significant effects of exposure on *root* (p=0.014) and *suck* (p=0.015) 8 such that unexposed animals reached criterion more quickly (Table 3). For *snout* the effect 9 approached significance (p=0.06). There was no main effect of birth weight on any neonatal 10 behavior (data not shown).

11

12 Insert Table 3

13

3.2.3 Modeling All Three Variables as Main Effects: Entering GA, birth weight, and exposure into the same model for each of the neonatal reflexes did not improve model fit and added nothing to the results, although exposure maintained its main effect on outcome for *root* (p=0.009) and *suck* (p=0.02) and approached significance for *snout* (p=0.054). GA maintained its independent effect on *visual follow far* (p=0.017).

19

20 **3.3 Modeling Time-to-Criterion: Interaction Models**

21 3.3.1 Gestational Age (GA), Birth Weight and Exposure Status

22 To evaluate whether exposure was associated with different effects at different levels of GA and/or

23 birth weight, and whether these covariates operated differently in exposed vs. unexposed infants, a

24 number of interaction terms were included in the models for *root, suck* and *snout*.

For each neonatal reflex interaction terms were included together with main effects in a number of models as follows: Model (a): a three-way interaction between birth weight, exposure and GA; and 3 two-way interaction models: Model (b) between exposure and GA; Model (c) between exposure and birth weight; and Model (d) between birth weight and GA. Models (a) to (d) were compared in terms of log likelihood statistics (-2LL), with those showing a lower -2LL, with at least one statistically significant model term being optimal.

7

For the *root* reflex, the optimal interaction model was Model (d) which included the two-way interaction between GA and birth weight (p=0.016), together with the main effects of each. This had the lowest -2LL statistic of all *root* models described above (see Table 4 for all model parameters and relevant output) and compared favorably to the Omnibus Model. In the optimal *root* model, all terms were significant indicating that exposure (p=0.003), GA (p=0.035) and birth weight (p=0.046) are all significant factors in the acquisition of the '*root*' reflex, when an interaction between GA and birth weight is allowed for.

15

16 Insert Table 4

17

For the *suck* reflex, two models fitted almost equally well: Model (a) a three-way interaction between GA, birth weight and exposure (p=0.012) and Model (c) a two-way interaction between exposure and birth weight (p=0.01). Both models compared favorably with, and were a significant improvement on, the Omnibus Model. The three-way model term of birth weight (p=0.038) and the three way interaction between birth weight, GA and exposure (p=0.024) were statistically significant with exposure maintaining an effect that approached significance (p=0.075) once its interaction with birth weight and GA was controlled. For the two-way model (which included exposure and birth

- 1 weight as the interaction term), both birth weight (p=0.035) and the interaction term (p=0.02) were 2 statistically significant, with exposure maintaining an effect that approached significant (p=0.055).
- 3

For the *snout* reflex, the optimal interaction model was Model (d) which included the two-way 4 5 interaction term between GA and birth weight together with the main effects of each. This had the 6 lowest -2LL statistic of all snout models and compared favorably to the Omnibus Model (Table 4). In 7 this model, although the interaction term was not in itself statistically significant, by controlling for 8 the possible interaction between GA and birth weight, its inclusion strengthened the overall model 9 and resulted in a significant effect of exposure (p=0.033). None of the remaining model terms GA, 10 birth weight, and the interaction between GA and birth weight, were statistically significant (p>0.05). 11 All interaction models reported here improved on model fit for models which included main effects 12 alone.

13

In summary, the interaction models indicated that although there was no main effect of GA or birth weight on any variable other than *visual follow far*, there were various interactions between GA, birth weight and exposure, that, when controlled, added strength to the models being evaluated and indicated important influences of birth weight and/or GA on the effect of exposure. In general, the data indicate that low birth weight and/or low GA exacerbates the effect of exposure, while increasing GA and/or birth weight mitigates the detrimental effect of exposure.

- 20
- 21
- 22

1 4. Discussion

2 This study demonstrates that the acquisition of three neonatal survival reflexes, root, suck and snout, 3 was significantly delayed in rhesus macaques receiving a single thimerosal (Th)-containing HB 4 vaccine at birth. All infants remained healthy for the duration of the study suggesting that there were 5 no health-related changes that may have affected the acquisition of reflexes. We sought, therefore, to 6 examine what other variables might either account for, or influence, these observations. In general, 7 as GA increased animals reached criterion earlier whereas animals of lower GA were relatively 8 delayed. This effect was only significant when exposure was taken into account. For exposed 9 animals, the effect of increasing GA was to mitigate the detrimental effect of exposure. Since there 10 was no linear or additive relationship between GA and birth weight in this study, these observations 11 cannot be readily accounted for by, for example, an effect of dose-of-exposure alone. It is plausible 12 that while reflex acquisition per se is not influenced by GA alone, the brain of the less mature neonate 13 may be more susceptible to neurotoxic injury manifesting as delayed reflex acquisition.

14

15 Although the basis for the effect of birth weight is not known, it is plausible that lower birth weight 16 infants are more susceptible to what may be a dose-dependent toxicity of Th or some other HB 17 vaccine constituent, such as aluminum. The effects on time-to-criterion appeared to be non-18 random: of the four survival reflexes, three were significantly negatively affected by exposure, while 19 for *startle*, the fourth survival reflex, effects were similar but did not reach statistical significance. 20 Interaction effects were observed with these same reflexes for both birth weight and GA. This 21 interaction involved mitigation of the effect of exposure in a way that is biologically plausible i.e. 22 reduced time-to-criterion with increasing GA and birth weight.

23

1 Neurodevelopmental tests are used for both human and non-human primate neonates to study 2 developmental status [32-36] and these tests typically involve assessment of reflexes and orienting 3 responses to visual and auditory stimuli as reported here. These test batteries are important in 4 assessing the potential neurodevelopmental toxicity of chemicals and drugs [37-40]. They provide a 5 broad-based evaluation of a range of nervous system functions at a period of life when learning and 6 adaptation are particularly critical. Non-human primates are an especially appropriate test species 7 because of their similarities to humans in complexity of brain function and prolonged intrauterine 8 brain development (reviewed by [41]).

9

10 Our study design does not enable us to determine whether it is the vaccine per se, the exposure to 11 Th, or a combination of both, that is causing the observed effects. None-the-less, the developing 12 brain is considered the most vulnerable organ to mercury exposure [42], and experimental studies 13 suggest that the brainstem - whose function is central to the reflexes described herein - may be one 14 of the more sensitive targets [43-44]. Dietary methylmercury has been shown to accumulate in the 15 brain of fish, resulting in histopathological damage, significantly reduced neural enzyme activity, and 16 altered behavior [45]. Pathological injury was observed to have started in the brainstem, extending to 17 other areas of the brain at higher exposure levels. In a mouse model, exposure to mercury vapor 18 resulted in a preferential accumulation of mercury in the brainstem, regardless of concentration used 19 [46]. Similarly, after intramuscular injection, inorganic mercury accumulated in brainstem motor 20 nuclei of mice [47]. In clinical studies of mercury poisoning, exposure to organic mercury either pre-21 or post-natally resulted in brainstem defects in children [48-52]. Since the acquisition of motor 22 reflexes is controlled by the brainstem, it is possible that very early exposure to ethyl mercury may 23 adversely affect emerging brainstem function [53]. Brainstem injury may then disturb the 24 development or functioning of higher structures [54-55, reviewed by 56].

1 We adjusted and standardized the Th concentration in order to give animals a clinically relevant 2 exposure dose and allow meaningful comparisons. As in clinical practice, however, the final dose in 3 terms of µg/kg body weight was dependent solely on the infants' birth weight. Stajich et al. [4] 4 examined blood mercury levels in US infants after receiving a single dose of HB vaccine and found 5 the highest levels of mercury in pre-term infants suggesting that newborns, especially pre-term 6 infants, may have decreased ability to eliminate mercury. Pichichero et al. [57] also measured blood 7 mercury in infants after receiving a single Th-containing HB vaccine at birth. Since blood samples 8 were only collected once from each infant and at time points ranging from 12 hours to 30 days post 9 exposure, it is not possible to draw any convincing conclusions from this study regarding the 10 disposition of mercury in the blood of newborns after a single HB vaccine. Despite this, however, 11 their data showed that in those infants for whom a blood sample was collected between 12 hours 12 to 2 days post exposure, blood mercury levels varied greatly between subjects at each time point 13 [57], suggesting that the ability to eliminate mercury varied among infants. Perhaps more 14 importantly, the birth weight of infants ranged from 2.3 to 4.5 kg - almost a 2-fold difference - and 15 yet data were not analyzed in respect of birth weight. Current pediatric immunization 16 recommendations are for primary HB vaccination at birth [1-2] with no precautions for premature 17 and/or low birth weight babies [1]. In fact the World Health Organization recommends that the 18 birth-dose HB vaccine be given to preterm infants but if their birth weight is <2000 g, the vaccine 19 dose at birth should not be counted towards the primary series, and three additional doses should be 20 administered [58]. Based upon the current findings in term, normal weight-range neonatal macaques, 21 it may be that premature and/or low birth weight neonates are at increased risk of neurotoxicity. We 22 have previously shown that very low birth weight and/or premature rhesus macaque infants display 23 much longer delays in acquiring these same reflexes [59]. It is notable that, within low birth weight 24 animals, males have significantly delayed development for some reflexes relative to females,

including the *suck* reflex (9). The acquisition of reflexes in infant primates of low birth weight
 and/or GA receiving a Th-containing HB vaccine at birth should be examined.

3

There have been several animal studies looking at the effects of thimerosal-containing vaccines 4 5 (TCVs) and/or Th on neurodevelopment, behavior, immune function, and toxicology [12, 60-63]. 6 Burbacher et al. [12] examined the disposition and distribution of mercury in the brain of 7 cynomolgus macaques administered methyl mercury or TCVs. In their study, cynomolgus macaques 8 received TCVs and were sacrificed at various time-points post vaccination. While their data 9 demonstrated that tissue distribution and clearance rates differed between methyl mercury and Th-10 exposed infants, the proportion of inorganic mercury in the brain was substantially higher for 11 animals receiving TCVs [12]. Once inorganic mercury has accessed the brain, its half-life is much 12 longer than both ethyl and methyl mercury, and it has the potential to accumulate in cases of 13 prolonged or repeated exposure (reviewed by Rooney, [64]). If, in our on-going investigations, Th is 14 found to be driving the detrimental development effects, a dose-response study would be warranted. 15 Our findings provide an important rationale for determining what factors in the HB vaccine may be 16 responsible for these clinical observations. This should also include aluminum hydroxide which is 17 used as an adjuvant in many vaccines, including the HB vaccine formulation used for this study. 18 Studies are underway to examine this, and the consequences of repeated and/or additional vaccine 19 exposures on the natural course of neurodevelopment.

20

1 5. Conclusions

2 In summary, this study provides preliminary evidence of abnormal early neurodevelopmental 3 responses in male infant rhesus macaques receiving a single dose of Th-containing HB vaccine at 4 birth and indicates that further investigation is merited. Birth weight and GA appear to be important 5 variables that might predicate susceptibility. This study design was not able to determine whether it 6 was the vaccine *per se*, the exposure to thimerosal, or a combination of both, that caused these 7 effects. While primate testing forms an important part of pre-clinical safety assessment of vaccines 8 intended for human use [65], the outcomes reported here are not included in the current CDC 9 recommendations for hepatitis B vaccine safety testing [66]. A replication study in a larger cohort of 10 infants is underway that extends these investigations to other areas of clinical concern such as 11 emerging cognition, long-term learning and behavior, and neuroimaging studies of brain structure 12 and function.

13

14 Acknowledgements

15 We thank Drs. Saverio Capuano and Mario Rodriguez for veterinary assistance; Amanda Dettmer, 16 Daniel Hollenbeck, Carrie Redinger, Dave McFarland, Melanie O'Malley and Megan Rufle for 17 technical support. We would like to express our gratitude to Robert Sawyer, Troy and Charlie Ball, and 18 Dr. Jeff Bradstreet. We are indebted to the late Gerald Ruppenthal who assisted in the study design, 19 training and implementation of the infant primate developmental measures prior to his death in 2005. 20 This work was supported by the Johnson Family, the late Liz Birt, SafeMinds, the Autism Research 21 Institute, The Ted Lindsay Foundation, the Greater Milwaukee Foundation, David and Cindy 22 Emminger, Sandy McInnis, Elsie Roberts and Vivienne McKelvey. LH and AJW designed the study

but were not involved in data collection and statistical analysis. LH was also responsible for coordinating all aspects of the study. LAH was responsible for newborn primate care and neurodevelopmental assessments. CS and GS were responsible for data analyses; JLT was responsible for ultrasounds of pregnant dams and newborn injections; and DA, LB and ERW were responsible for the production of TCVs.

6

7 Conflict of Interest Statement

8 Prior to 2005, CS and AJW acted as paid experts in MMR-related litigation on behalf of the plaintiff.
9 LH has a child who is a petitioner in the National Vaccine Injury Compensation Program. For this
10 reason, LH was not involved in any data collection or statistical analyses to preclude the possibility of
11 a perceived conflict of interest.

Correction of the second secon

- 12
- 13
- 14

1 References

2	1.	Hepatitis B: A comprehensive strategy for eliminating transmission in the United States
3		through universal childhood vaccination: Recommendations of the Immunization Practices
4		Advisory Committee (ACIP), CDC Morbidity and Mortality Weekly Report 40 (1991) RR13,
5		1-19.
6	2.	Universal Hepatitis B Immunization, Committee on Infectious Diseases, Pediatrics 89 (1992)
7		795-800.
8	3.	Dórea, J.G., Marques, R.C., Brandão, K.G. Neonate Exposure to Thimerosal Mercury from
9		Hepatitis B Vaccines. Am. J. Perinatol. (2009) 26, 523-527. Mar 12. [Epub ahead of print]
10	4.	Stajich, G.V., Lopez, G.P., Harry, S.W., Sexson, W.R., Iatrogenic exposure to mercury after
11		hepatitis B vaccination in preterm infants, J. Pediatr. 136 (2000) 679-81.
12	5.	Ono, H., Sakamoto, A., Sakura, N., Plasma total glutathione concentrations in healthy
13		pediatric and adult subjects, Clin. Chim. Acta 312 (2002) 227-229.
14	6.	Erden-Inal, M., Sunal, E., Kanbak, G., Age-related changes in the glutathione redox system,
15		Cell. Biochem. Funct. 20 (2002) 61-66.
16	7.	Ruppenthal, G.C., Sackett, G.P., Research Protocol and Technician's Manual. (2nd ed.):
17		Infant Primate Research Laboratory, University of Washington, 1992.
18	8.	Schneider, M.L., Suomi, S.J., Neurobehavioral assessment in rhesus monkey neonates
19		(Macaca mulatta): developmental changes, behavioral stability, and early experience, Infant
20		Behav. Dev. 15 (1992) 155-177.

1	9.	Kroeker, R., Sackett, G., Reynolds, J., Statistical methods for describing developmental
2		milestones with censored data: effects of birth weight status and sex in neonatal pigtailed
3		macaques, Am. J. Primatol. 69 (2007) 1313-1324.
4	10.	Satoh, H., Yasuda, N., Shimai, S., Development of reflexes in neonatal mice prenatally
5		exposed to methylmercury and selenite, Toxicol. Lett. 25 (1985) 199-203.
6	11.	Burbacher, T.M., Sackett, G.P., Mottet, N.K., Methylmercury effects on the social behavior
7		of Macaca fascicularis infants, Neurotoxicol Teratol. 12 (1990) 65-71.
8	12.	Burbacher, T.M., Shen, D.D., Liberato, N., Grant, K.S., Cernichiari, E., Clarkson, T.,
9		Comparison of blood and brain mercury levels in infant monkeys exposed to methyl
10		mercury or vaccines containing thimerosal, Environ. Health Perspect. 113 (2005) 1015-1021.
11	13.	Gunderson, V.M., Grant, K.S., Burbacher, T.M., Fagan, .JF. 3rd, Mottet, N.K., The effect of
12		low-level prenatal methylmercury exposure on visual recognition memory in infant crab-
13		eating macaques, Child Dev. 57 (1986) 1076-83.
14	14.	Gunderson, V.M., Grant-Webster, K.S., Burbacher, T.M., Mottet, N.K., Visual recognition
15		memory deficits in methylmercury-exposed Macaca fascicularis infants, Neurotoxicol. Teratol.
16		(1988) 10, 373-379.
17	15.	Rice, D.C., Gilbert, S.G., Early chronic low-level methyl mercury poisoning in monkeys
18		impairs spatial vision, Science 216 (1982) 759-761.
19	16.	Rice, D.C., Gilbert, S.G., Effects of developmental exposure to methyl mercury on spatial
20		and temporal visual function in monkeys, Toxicol. Appl. Pharmacol. 102 (1990) 151-163.
21	17.	Rossi, A.D., Ahlbom, E., Ogren, S.O., Nicotera, P., Ceccatelli, S., Prenatal exposure to
22		methylmercury alters locomotor activity of male but not female rats, Exp. Brain Res. 117
23		(1997) 428–436.

1	18.	Sakamoto, M., Wakabayashi, K., Kakita, A., Takahashi, H., Adachi, T., Nakano, A.,
2		Widespread neuronal degeneration in rats following oral administration of methylmercury
3		during the postnatal developing phase: a model of fetal-type Minamata disease, Brain Res.
4		784 (1988) 351-354.
5	19.	White, J.F., Thimerosal: Chemistry, Toxicology and Effects on Biological Systems. In:
6		Autism Research Challenges, ed. L.B. Zhao, Nova Science Publishers, Inc. Chapter 7, pp.
7		137-167, 2007.
8	20.	Branch, D.R, Gender-selective toxicity of thimerosal, Exp. Toxicol. Pathol. 61 (2009) 133-
9		136.
10	21.	Malagutti, K.S., da Silva, A.P., Braga, H.C., Mitozo, P.A., Soares dos Santos, A.R., Dafre, A.
11		L., de Bem, A. F., Farina, M., 17β-estradiol decreases methylmercury-induced neurotoxicity
12		in male mice, Environ. Toxicol. Pharmacol. 27 (2009) 293-297.
13	22	Gao, C.H., Yan, Y., Tian, Y., Wang, H.F., Xie, X., Zhou, X.D., Yu, X.G., Yu, S., Tong,
14		Q.X., Zhou, Shen, X.M., Prenatal exposure to mercury and neurobehavioral development of
15		neonates in Zhoushan city, China, Environ. Res. 105 (2007), 390–399.
16	23.	Sackett, G.P., Ruppenthal, G.C., Davis, A.E., Survival, Growth, Health, and Reproduction
17		Following Nursery Rearing Compared with Mother Rearing in Pigtailed Monkeys (Macaca
18		nemestrina), Am. J. Primatol., 56 (2002) 165-184.
19	24.	Suomi, S.J., Mineka, S., DeLizio, R.D., Short- and long-term effects of repetitive mother-
20		infant separations on social development in rhesus monkeys. Dev. Psychol. 19 (1983) 770-
21		786.

1	25. Ruppenthal, G., Sackett, G., Nursery care of at-risk nonhuman primates, In: Nursery rearing
2	of nonhuman primates in the 21st century, ed. G. Ruppenthal, G. Sackett and K. Elias,
3	University of Chicago, Chicago IL, Chapter 18, pp 371-390, 2006.
4	26. Ruppenthal, G., Weight gain and intake requirements in nursery-reared macaques. Proc. 4th
5	Ann. Dr. Scholl Conference on the Nutrition of Captive Wild Animals, Lincoln Park
6	Zoological Society, Chicago, 1989.
7	27. Brazleton, T.B, The Brazleton Neonatal Behavior Assessment Scale: Introduction. Monogr.
8	Soc. Res. Child Dev. 43 (1978) 1-13.
9	28. Chamove, A.S., Molinaro, T.J., Monkey retardate learning analysis, J. Ment. Defic. Res. 22
10	(1978) 37-48.
11	29. Ruppenthal, G.C., Walker, G.C., Sackett, G.P., Rearing infant monkeys (Macaca nemestrina) in
12	pairs produces deficient social development compared with rearing in single cages, Am. J.
13	Primatol. 25 (1991) 103-113.
14	30. Sackett, G., Ruppenthal, G., Hewitson, L., Simerly, C., Schatten, G., Neonatal behavior and
15	infant cognitive development in rhesus macaques produced by assisted reproductive
16	technologies, Dev. Psychobiol. 48 (2006) 243-265.
17	31. Cox, D. R., Regression Models and Life Tables (with Discussion). J. Royal Stat. Soc., Series
18	B 34 (1972) 187-220.
19	32. Abrams, S.M., Field, T., Scafidi, F. and Prodromidis, M., Newborns of depressed mothers,
20	Infant Mental Health J. 16 (1995) 231–235.

1	33. Amiel-Tison, C., Barrier, G., Schnider, S.M., Levinson, G., Hughes, S.C., Stefani, S.J., A new
2	neurologic and adaptive capacity scoring system for evaluating obstetric medications in full-
3	term newborns, Anesthesiology 56 (1982) 340-350.
4	34. Karmel, B.Z., Gardner, J.M., Freedland, R.L., Neonatal neurobehavioral assessment Bayley I
5	and II scores of CNS-injured and cocaine-exposed infants, Ann. N.Y. Acad. Sci. 846 (1998)
6	391-395.
7	35. Golub, M.S., Gershwin, M.E., Standardized neonatal assessment in the rhesus monkey. In:
8	Research in Perinatal Medicine, ed. P.W. Nathanielsz and J.T. Parer, Perinatology Press,
9	Ithaca, NY, 1984.
10	36. Rogers, W.R. Behavioral Testing. In: Non-human primates in perinatal research. Brans,
11	Y.W., Keuhl, T.J., eds. New York: John Wiley and Sons: 411-420, 1988.
12	37. Laughlin, N.K., Lasky, R.E., Giles, N.L., Luck, M.L., Lead effects on neurobehavioral
13	development in the neonatal rhesus monkey (Macaca mulatta), Neurotoxicol. Teratol. 21
14	(1999) 627-638.
15	38. He, N., Bai, J., Champoux, M., Suomi, S.J., Lidow, M.S., Neurobehavioral deficits in
16	neonatal rhesus monkeys exposed to cocaine in utero, Neurotoxicol. Teratol. 26 (2004) 13-21.
17	39. Stahlmann, N., Härtel, C., Knopp, A., Gehring, B., Kiecksee, H., Thyen, U., Predictive value
18	of neurodevelopmental assessment versus evaluation of general movements for motor
19	outcome in preterm infants with birth weights <1500 g, Neuropediatrics 38 (2007) 91-99.
20	40. Medoff-Cooper, B., Shults, J., Kaplan, J., Sucking behavior of preterm neonates as a
21	predictor of developmental outcomes, J. Dev. Behav. Pediatr. 30 (2009) 16-22.

1	41. Golub, M.S., Use of monkey neonatal neurobehavioral test batteries in safety testing
2	protocols, Neurotoxicol. Teratol.12 (1990) 537-541.
3	42. Grandjean, P. and Perez, M., Developmental neurotoxicity: implications of methylmercury
4	research, Intl. J. Environ. Health 2 (2008) 417 -428.
5	43. Sakamoto, M., Nakano, A., Akagi, H., Declining Minamata male birth ratio associated with
6	increased male fetal death due to heavy methylmercury pollution, Environ. Res. 87 (2001)
7	92-98.
8	44. Grandjean, P., Murata, K., Budtz-Jørgensen, E., Weihe, P., The brainstem as a target of
9	developmental methylmercury toxicity. 7th International Conference on Mercury as a Global
10	Pollutant Ljubljana, Slovenia, 2004.
11	45. Berntssen, M.H., Aatland, A., Handy, R.D., Chronic dietary mercury exposure causes
12	oxidative stress, brain lesions, and altered behaviour in Atlantic salmon (Salmo salar) parr,
13	Aquat. Toxicol. 65 (2003) 55-72.
14	46. Warfvinge, K., Mercury distribution in the mouse brain after mercury vapor exposure. Int. J.
15	Exp. Pathol. 76 (1995) 29-35.
16	47. Arvidson, B., Accumulation of inorganic mercury in lower motor neurons of mice,
17	Neurotoxicology 13 (1992) 277-280.
18	48. Amin-Zaki, L., Majeed, M.A., Clarkson, T.W., Greenwood, M.R., Methylmercury poisoning
19	in Iraqi children: clinical observations over two years, Br. Med. J. 1 (1978) 613-616.
20	49. Magos, L., Brown, A.W., Sparrow, S., Bailey E., Snowden, R.T., Skipp, W.R., The
21	comparative toxicology of ethyl-and methylmercury, Arch. Toxicol. 57 (1985) 260-267.

1	50. Counter, S.A., Neurophysiological anomalies in brainstem responses of mercury-exposed
2	children of Andean gold miners, J. Occup. Environ. Med. 45 (2003) 87-95.
3	51. Murata, K., Weihe, P., Budtz-Jørgensen, E., Jørgensen, P.J., Grandjean, P., Delayed
4	brainstem auditory evoked potential latencies in 14-year-old children exposed to
5	methylmercury, J. Pediatrics 144 (2004) 177-183.
6	52. Bernard, S., Enayati, A., Redwood, L., Roger, H., Binstock, T., Autism: a novel form of
7	mercury poisoning, Med. Hypotheses 56 (2005) 462-471.
8	53. Wakefield, A.J., Stott, C., Lopresti, B.J., Tomko, J., Houser, L., Sackett, G., Hewitson, L.,
9	Pediatric vaccines influence primate behavior, and brain stem volume and opioid ligand
10	binding, 4th Annual International Meeting for Autism Research (IMFAR), London, UK,
11	2008.
12	54. Geva, R., Feldman, R., A neurobiological model for the effects of early brainstem
13	functioning on the development of behavior and emotion regulation in infants: implications
14	for prenatal and perinatal risk, J. Child Psychol. Psychiatry 49 (2008) 1031-41.
15	55. Tanguay, P.E., Edwards, R.M., Electrophysiological studies of autism: the whisper of the
16	bang, J. Autism Dev. Disord.12 (1982) 177-184.
17	56. McGinnis, W.R., Miller, V.M., Audhya, T., Edelson, S., Neurotoxic brainstem impairment as
18	proposed threshold event in autistic regression. Taylor & Francis, CRC Press, 2009 (In
19	Press).
20	57. Pichichero, M.E., Gentile, A., Giglio, N., Umido, V., Clarkson, T., Cernichiari, E., Zareba,
21	G., Gotelli, C., Gotelli, M., Yan, L., Treanor, J., Mercury levels in newborns and infants after
22	receipt of thimerosal-containing vaccines, Pediatrics 121 (2008) 208-214.

1	58. World Health Organization: Weekly Epidemiological Record (WER), 79 (2004) 263-274.
2	59. Dettmer, A.M., Houser, L.A., Ruppenthal, G.C., Capuano, S., Hewitson, L., Growth and
3	developmental outcomes of three high-risk infant rhesus macaques (Macaca mulatta), Am. J.
4	Primatol. 69 (2007) 503-518.
5	60. Hornig, M., Chian, D., Lipkin, W.I., Neurotoxic effects of postnatal thimerosal are mouse
6	strain dependent, Mol. Psychiatry 9 (2004) 833-845.
7	61. Havarinasab, S., Haggqvist, B., Bjorn, E., Pollard, K.M., Hultman, P., Immunosuppressive
8	and autoimmune effects of thimerosal in mice, Toxicol. Appl. Pharmacol. 204 (2005) 109-
9	121.
10	62. Berman, R.F., Pessah, I.N., Mouton, P.R., Mav, D., Harry, J. Low-level neonatal thimerosal
11	exposure: further evaluation of altered neurotoxic potential in SJL mice, Toxicol. Sci. 101
12	(2008) 294-309.
13	63. Minami, T., Miyata, E., Sakamoto, Y., Yamazaki, H., Ichida, S, Induction of metallothionein
14	in mouse cerebellum and cerebrum with low-dose thimerosal injection, Cell Biol. Toxicol.
15	Apr 9 2009 [Epub ahead of print].
16	64. Rooney, J.P., The role of thiols, dithiols, nutritional factors and interacting ligands in the
17	toxicology of mercury, Toxicology 234 (2007) 145-156.
18	65. Kennedy, R.C., Shearer, M.H., Hildebrand, W., Nonhuman primate models to evaluate
19	vaccine safety and immunogenicity, Vaccine 15 (1997) 903-908.
20	66. Current trends hepatitis B virus vaccine safety: Report of an inter-agency group, Morbidity
21	and Mortality Weekly Report 31 (1982) 465-467.

1

Service of the servic

1 Table Captions

2

3 Table 1. Neonatal Reflexes Measured, Rating Categories, and Criterion Responses.

4

5 Table 2. Neonatal Reflexes: Mean Time-to-Criterion with log-rank statistic and associated p 6 values, for exposed (E) and unexposed (U) groups. Kaplan-Meier survival analysis was used to 7 compare age in days at reaching neonatal basic motor reflexes and sensorimotor responses. The log 8 rank test identified statistically significant differences ($p \le 0.05$) for the achievement of these 9 milestones between exposed and unexposed groups for three basic motor reflexes (root, suck and 10 snout). There were no statistically significant differences between the groups in time-to-criterion for 11 the remaining neonatal behaviors. *n=6 for *snout* (scored as missing data for one infant; **censored 12 at Day 14 (one infant in the exposed group did not reach criterion during the 14 day testing period 13 for this study).

14

15 Table 3. Main Effects Models for Exposure, Gestational Age (GA) and Birth Weight: 16 Significant Predictors. For each neonatal behavior Model, '-2LL' gives the value of the model 17 when all variables are included. Better models have lower values for -2LL. The change from the 18 Omnibus Model (which assumes all effects are 0) to the specified model containing all variables, is 19 represented by the χ^2 and its associated p value. P < 0.05 indicates an improved model over the 20 Omnibus. For Model terms the $Exp(\beta)$ or Hazard Ratio indicates the rise in risk of outcome 21 (achievement of criterion) associated with a one unit rise in the predictor after controlling for all 22 other terms in the model. Its associated value indicates the probability of an $Exp(\beta)$ of this 23 magnitude being generated by chance alone. For all analyses, Exposure=0 is the Indicator variable.

The exposure effect is therefore the effect of being unexposed. In other words for the *root* reflex,
 unexposure is associated with a 4.84 risk of meeting criterion for unexposed relative to an exposed
 animal. Other effects are of a rise in the predictor associated with a rise in risk hazard.

Table 4. Best Fit Interaction Models for Root and Suck Reflexes. For each neonatal behavior Model, '-2LL' gives the value of the model when all variables are included. Better models have a lower -2LL. The change from the Omnibus Model (which assumes all effects are 0) to the specified model containing all variables, is represented by the χ^2 and its associated *p* value. P < 0.05 indicates an improved model over the Omnibus. For Model terms the $Exp(\beta)$ indicates the rise in risk of outcome (achievement of criterion) associated with a one unit rise in the predictor, after controlling for all other terms in the model. Its associated value indicates the probability of an $Exp(\beta)$ of this magnitude being generated by chance alone. For all analyses, Exposure=0 is the Indicator variable. While it is important to include Main Effects when modeling an interaction term, interpretation of any associated parameter estimates should be avoided when the interaction term is statistically significant. Parameter estimates for Main Effects are included here for reference and completeness.

1 Figure Legends

2 Figure 1. The Kaplan-Meier (K-M) survival curve is a step-function used here to indicate the 3 estimated proportion of animals having reached criterion at various time points from the start of the 4 study period. K-M curves for neonatal reflexes in exposed (green) and unexposed (blue) rhesus 5 macaque infants are presented: reflexes for time-to-criterion were measured daily from birth through 6 Day 14. Significant differences were observed in the mean age (days) at reaching criterion for root (A; 7 p=0.004), suck (B; p=0.002) and snout (C; p=0.03). The value on the y axis represents the proportion 8 of animals not reaching criterion by the time (in days) represented on the x-axis. Any point on the 9 curve gives the proportion of infants still not having reached criterion at a particular time after the 10 start of the observation period. For example, in A (root) 58% of the unexposed animals remained at 11 Day 1 (meaning that 42% had reached criterion at this stage compared to around 9% of the exposed 12 animals).

13

1 Table 1

Test Reflex	Rating Category/Definition	Criterion Score	
Rooting (left, right)	0 = no rooting 1 = partial rooting, doesn't move completely to object 2 = weak slow and/or intermittent move to object 3 = strong, quick vigorous move to object	3	
Snouting	0 = none 1 = partial response, mouth not open 2 = weak, slow response (opens mouth) but sluggish 3 = strong, quick full response (opens mouth)	3	
Sucking	 0 = none 1 = partial response, mostly mouthing, little sucking 2 = weak sucking 3 = strong suck but biting first 	3	
Auditory startle	0 = none 1 = whole body jerk	1	
Grasping (left, right; hands, feet)	0 = no grasp 1 = partial 2 = weak 3 = strong	3	
Clasping	0 = no clasp 1 = loose clasp 2 = firm clasp 3 = climbs off	2	
Auditory orient (left, right)	0 = none 1 = partial 2 = full orient	2	
Visual orient (near, far)	0 = none 1 = head moves 2 = brief contact 3 = prolonged contact	3	
Visual follow (left, right; near, far)	0 = no contact 1 = contact, no follow 2 = incomplete follow 3 = complete follow	3	

2

3

Table 2 1

	Mean Time-to-Criterion (Days)				Kaplan-Meier Survival (Log Rank)			
Reflex	Exposed (n=13)**		Unexposed (n=7)*		Chi Square	1		
	Mean	95% CI	Mean	95% CI	(χ^2)	<i>p</i> value		
Survival Reflexes								
Root	2.33	1.80 - 2.85	1.11	0.61 – 1.61	8.19	0.004		
Suck	3.38	2.42 - 4.35	1.50	0.90 - 2.10	9.52	0.002		
Snout*	3.46	2.08 - 4.85	1.58	0.89 – 2.27	4.58	0.032		
Auditory Startle**	2.92	0.94 - 4.90	0.93	0.39 – 1.47	2.51	0.113		
Motor Reflexes					·			
Grasp Hand	0.85	0.57 – 1.13	0.50	0.50 - 0.50	3.41	0.065		
Grasp Foot	0.77	0.56 – 0.98	0.7	0.26 - 1.24	0.36	0.956		
Clasp Hand	1.37	0.79 – 1.94	1.21	0.38 - 2.05	0.11	0.742		
Clasp Foot	1.65	0.97 – 2.34	1.29	0.59 – 1.98	0.69	0.407		
Sensorimotor Reflexes	6							
Auditory Orient	3.21	1.89 - 3.77	2.83	0.65 - 5.78	0.32	0.570		
Visual Orient (Nr)	1.77	0.96 - 2.58	2.36	0.73 – 3.98	0.18	0.670		
Visual Orient (Far)	1.73	0.92 – 2.54	2.36	0.73 – 3.40	0.14	0.712		
Visual Following (Nr)	3.29	2.35 - 4.23	3.04	1.36 - 4.71	0.03	0.864		
Visual Following (Far)	2.91	1.48 - 4.33	3.68	1.87 – 5.49	0.88	0.348		

2 3 CI, confidence interval; Nr, near.

1 Table 3

2

Reflex	Variables in Model	Model -2LL	Change Omnibus to Model		Model Effects	Exp (β)	95% CI		<i>p</i> Exp (β)
			χ^2	$p(\chi^2)$			Lower	Upper	
	Exposure	82.74	6.90	0.03	Exposure	4.841	1.481	15.821	0.009
	GA	02.74	0.90	0.03	Gestational Age	1.046	0.966	1.133	0.264
_	Exposure	83.67	5.97	0.05	Exposure	4.154	1.330	12.973	0.014
Root	Birth Weight	65.07	5.97	0.03	Birth Weight	1.002	0.996	1.007	0.555
	Exposure				Exposure	4.844	1.479	15.862	0.009
	GÂ	82.74	6.90	0.07	Gestational Age	1.047	0.953	1.149	0.338
	Birth Weight				Birth Weight	1.000	0.993	1.007	0.986
	Exposure	00.00		0.026	Exposure	4.411	1.337	14.559	0.015
	GÂ	82.89	6.66	0.036	Gestational Age	0.979	0.902	1.062	0.609
	Exposure		7.65	0.022	Exposure	4.282	1.323	13.864	0.015
Suck	Birth Weight	81.89			Birth Weight	0.996	0.990	1.003	0.282
	Exposure GA Birth Weight	81.75	7.80	0.05	Exposure	4.128	1.251	13.624	0.020
					Gestational Age	0.984	0.907	1.068	0.703
					Birth Weight	0.996	0.990	1.003	0.304
	Exposure GA	77.70	8.31	0.016	Exposure	0.786	0.289	2.134	0.636
					Gestational Age	1.155	1.031	1.293	0.013
Visual	Exposure Birth Weight	83.90	2.11	0.348	Exposure	0.689	0.264	1.800	0.447
Follow Far					Birth Weight	1.004	0.997	1.011	0.255
1 11	Exposure GA Birth Weight	76.97	9.04	0.03	Exposure	0.859	0.310	2.381	0.769
					Gestational Age	1.003	0.996	1.012	0.390
					Birth Weight	1.158	1.027	1.306	0.017
	Exposure		3.84	0.15	Exposure	3.246	0.973	10.827	0.055
	GÂ	77.61		0.15	Gestational Age	1.026	0.941	1.118	0.560
	Exposure	77.95	3.50	0.17	Exposure	3.171	0.949	10.600	0.061
Snout	Birth Weight		5.50	0.17	Birth Weight	1.000	0.994	1.006	0.930
	Exposure				Exposure	3.270	0.982	10.894	0.054
	GÂ	t 77.33	4.12	0.25	Gestational Age	1.043	0.938	1.160	0.437
	Birth Weight				Birth Weight	0.998	0.991	1.005	0.601

Table 4

Reflex	Variables in Model	Model -2LL	Change Omnibus to Model		Exp (β)	95% CI		<i>p</i> Exp(β)
			χ^2	$p(\chi^2)$		Lower	Upper	1
Root	Exposure	77.14	12.22	0.016	6.669	1.886	23.575	0.003
	GA				2.689	1.074	6.734	0.035
	Birth Weight				1.365	1.005	1.853	0.046
	GA*Birth Weight				0.998	0.996	1.000	0.046
Suck (Model a)	Exposure	76.78	12.76	0.012	0.004	0.000	1.746	0.075
	GA				0.936	0.856	1.025	0.153
	Birth Weight				0.990	0.981	0.999	0.038
	Exposure*GA*Birth Weight				1.000	1.000	1.000	0.024
Suck (Model b)	Exposure	76.35	13.19	0.01	0.001	0.000	1.173	0.055
	GÁ				0.947	0.870	1.031	0.211
	Birth Weight				0.990	0.980	0.999	0.035
	Exposure*Birth Weight				1.017	1.003	1.031	0.020
Snout	Exposure	75.46	5.99	0.20	3.766	1.114	12.73	0.033
	GÁ				1.178	0.911	1.524	0.210
	Birth Weight				1.697	0.796	3.616	0.171
	GA*Birth Weight				0.999	0.997	1.001	0.205
2 3								

